

Vital Surveillances

Genomic Insights into the 2024 HAdV-14p1 Strain in China: Traced to Early Prevalent Strains

Yali Jin¹; Jiangxia Wang²; Naiying Mao¹; Chunyu Zhu¹; Haiqing Zhou¹; Weiyan He¹; Hao Ding¹; Baicheng Xia¹; Aili Cui¹; Yan Zhang¹; Zhen Zhu^{1,†}; Hongmei Xu^{2,†}

ABSTRACT

Introduction: Human adenovirus type 14 variant p1 (HAdV-14p1) has been associated with severe respiratory infections worldwide. However, no cases of HAdV-14p1 infection have been reported in China since 2019. HAdV-14 was unexpectedly identified in 2024 in a pediatric patient hospitalized with bronchopneumonia in Chongqing, China. This study aims to elucidate the genetic characteristics of this strain and determine its phylogenetic relationship with previously circulating domestic and international strains.

Methods: Whole-genome sequencing was performed on the HAdV-14 strain Chongqing2024-115, followed by comprehensive phylogenetic and genetic variation analyses using all publicly available global HAdV-14 genome sequences from the GenBank database.

Results: Genome-based phylogenetic analysis classified global HAdV-14 strains into two well-supported clades: the HAdV-14 prototype clade and the variant HAdV-14p1 clade. The latter was further subdivided into three distinct subclades (I–III). Chongqing2024-115 clustered closely with earlier HAdV-14p1 subclade III and isolates from Beijing (2012) and Gansu (2013), showing high sequence identity (99.7%–99.9%). Compared with the genome of the HAdV-14 prototype strain (de Wit, AY803294), all three HAdV-14p1 subclades shared 99 specific nucleotide variations, including a six-nucleotide deletion in the knob domain of the fiber gene (nt751–756). The identification of subclade-specific variations and mutations unique to strain Chongqing2024-115 indicates ongoing microevolution within the HAdV-14p1 clade.

Conclusions: Our results confirm that the contemporary HAdV-14p1 strain shares a common genetic ancestry with earlier Chinese subclade III strains. Considering the association between HAdV-

14p1 and severe disease, sustained surveillance is essential to understand better its prevalence, evolution, and impact on public health in China.

Human adenoviruses (HAdVs) are non-enveloped, double-stranded DNA viruses with a genome size of approximately 35–36 kb (1). To date, 117 types have been identified and classified into seven species (A–G; <http://hadvwg.gmu.edu/>). Due to their diverse tissue tropisms, HAdV infections are associated with various illnesses, including acute respiratory infections, gastroenteritis, and conjunctivitis (2). Respiratory infections are frequently associated with specific species, including B (HAdV-3, 7, 14, 21, and 55), C (HAdV-1, 2, 5, and 6), and E (HAdV-4) (3).

HAdV type 14 (HAdV-14) was first identified in 1955 in a military recruit with acute respiratory disease (ARD) in the Netherlands, with only limited reports documented over subsequent decades (4). Approximately 50 years later, a novel and highly transmissible variant, designated human adenovirus type 14 variant p1 (HAdV-14p1), emerged and triggered multiple outbreaks in several countries, including the United States, the United Kingdom, Ireland, Canada, and Japan (5–9). These outbreaks affected both military and civilian populations and were associated with severe and fatal ARD cases (10). In China, HAdV-14p1 was initially detected in Guangzhou in 2010, with subsequent outbreaks in Beijing (2012), Liaoning (2012 and 2016), Gansu (2013), and Jiangsu (2015), resulting in substantial hospitalizations (11–15). HAdV-14p1 was sporadically detected in other provinces between 2011 and 2019. However, since 2019, no further cases have been reported in China.

Following the COVID-19 pandemic, HAdV-14 was newly identified in 2024 through sentinel surveillance in a pediatric patient hospitalized with

bronchopneumonia in Chongqing, China. To elucidate the genetic characteristics of this re-emerging HAdV-14 strain and determine its phylogenetic relationship with previously circulating domestic and international strains, we sequenced its complete genome; and performed comprehensive phylogenetic and genetic variation analyses using publicly available global HAdV-14 genome data from GenBank.

METHODS

Strain Source and Clinical Information

Sentinel surveillance for acute respiratory infections conducted since 2019 and covering 15 provinces did not detect HAdV-14 until 2024. In 2024, the HAdV-14 strain Chongqing2024-115 was isolated from a sputum sample collected from a 4-year-old male patient hospitalized with acute bronchitis in Chongqing, China. The patient presented with primary symptoms of fever, cough, and expectoration and required a 10-day hospitalization. Chest imaging revealed no significant abnormalities in either lung. Initial pathogen screening was positive for HAdV, and subsequent sequencing and analysis of the penton base, hexon, and fiber genes confirmed a HAdV-14 infection.

Genomic Sequencing and Annotation

Viral DNA was extracted from Chongqing2024-115 using a QIAamp DNA Mini Kit (Qiagen, Hilden, Germany). Whole-genome sequencing (WGS) was performed by iGeneTech Biotechnology Ltd. using a probe-based hybrid capture approach, followed by next-generation sequencing. This process yielded over 1 Gb of sequencing data, achieving complete genome coverage (100%) at a depth exceeding 8,000×. Sanger sequencing was used to validate genomic regions containing ambiguous bases. The genome was annotated using Geneious Prime software by mapping the sequences to the HAdV-14 prototype strain (de Wit, GenBank accession number AY803294). Complete genomic sequence of the Chongqing2024-115 strain in this study was deposited in the China National Microbiology Data Center with accession number NMDCN0009AD2.

Dataset

All available HAdV-14 WGSs were retrieved from GenBank. After removing incomplete and redundant sequences, a dataset of 67 WGSs from nine countries

(1955–2024) was established. This dataset included five Chinese strains (2010–2013) and 62 strains from eight countries and regions (1955–2024), including the United States, Canada, Japan, Honduras, Ireland, Nepal, the Netherlands, and Scotland. With the addition of the newly sequenced strain Chongqing2024-115 from this study, a final dataset of 68 WGSs was compiled for subsequent phylogenetic and genetic variation analyses.

Bioinformatics Analyses

Sequence alignment was performed using ClustalW in MEGA v7 (<http://www.megasoftware.net>), followed by pairwise similarity assessment using BioEdit v7.0.4.1 (<http://www.mbio.ncsu.edu/BioEdit/bioedit.html>). A maximum likelihood phylogenetic tree was constructed using MEGA v7, with bootstrap support (>80%) indicated at the corresponding nodes. In addition, a phylogenetic network based on WGSs was constructed using SplitsTree v4.19.2 (<http://www.splitsree.org>) for viral evolutionary analysis. Genetic variations were identified using the Snipit package, focusing on nucleotide mutations at a frequency of 100%. Recombination analysis was conducted using RDP4 (<http://web.cbio.uct.ac.za/~darren/rdp.html>) and SimPlot v3.5.1 (<http://sray.med.som.jhmi.edu/RaySoft/SimPlot>) (window size, 2,000 bp; step size, 100 bp).

RESULTS

Genomic Characterization

The complete genome of Chongqing2024-115 was 34,766 bp in length, with a GC content of 48.85%, consistent with the previously reported HAdV-14 genome (11). Annotation using the HAdV-14 prototype strain (AY803294) as a reference identified 38 protein-coding regions (Supplementary Table S1, available at <https://weekly.chinacdc.cn/>). Nucleotide sequence alignment revealed 99.6% identity with the prototype strain. Further analysis of 12 key functional regions, including genes encoding major capsid proteins (penton base, hexon, and fiber), core proteins (pV, pVII, pTP, and pIVa2), minor proteins (pIX, pIIIA, pVI, and pVIII), and the non-structural protein pX, showed that, with the exception of the fiber gene region (nt, 99.1%; aa, 98.7%), the remaining 11 regions exhibited high sequence similarity compared with the prototype strain (nt, 99.6%–100.0%; aa, 99.0%–100.0%) (Table 1).

TABLE 1. Sequence identity between strain Chongqing2024-115 and the HAdV-14 prototype strain (de Wit, AY803294).

Coding regions		Identity %	
		NT	AA
Major capsid protein	penton base	99.8	99.6
	hexon	99.8	99.8
	fiber	99.1	98.7
Core protein	pV	99.9	100.0
	PVII	99.8	100.0
	pTP	99.6	99.0
	pIVa2	99.7	99.2
Minor protein	pIX	100.0	100.0
	pIIIa	99.8	100.0
	pVI	99.8	99.5
	pVIII	99.8	100.0
Non-structural protein	pX	100.0	100.0
Whole genome sequence		99.6	

Abbreviation: NT=Nucleotide; AA=Amino Acid.

Phylogenetic Analysis

To elucidate the genetic relationship between Chongqing2024-115 and previously reported domestic and international HAdV-14 strains, phylogenetic analysis was conducted using the constructed WGS dataset (Figure 1). Consistent with previous findings, all 68 strains clustered into two well-supported clades: the HAdV-14 clade, comprising the prototype strain, and the distinct HAdV-14p1 clade (14). Notably, the HAdV-14p1 clade could be further subdivided into three subclades (I–III; bootstrap values >80%), a finding corroborated by phylogenetic network analysis, which revealed three independent evolutionary trajectories among these strains (Supplementary Figure S1, available at <https://weekly.chinacdc.cn/>). Nevertheless, all three subclades exhibited high sequence identity (99.5%–99.6%) with the HAdV-14 prototype strain, indicating a highly conserved HAdV-14 genome.

Chongqing2024-115 was phylogenetically assigned to HAdV-14p1 subclade III, which also included two other Chinese strains isolated from Beijing (2012) and Gansu (2013) (intra-subclade genetic distance, 0.00065). In contrast, subclades I and II exhibited broader geographical and temporal distributions. Subclade I comprised 54 strains from six countries, including the United States, Canada, Honduras, Ireland, Nepal, and Scotland (2003–2020; intra-subclade genetic distance, 0.00005). Subclade II consisted of three Chinese strains from Guangzhou

(2010) and Beijing (2011), along with five strains from the United States and Japan (2006–2024; intra-subclade genetic distance, 0.00008). High sequence identity was observed among the three subclades (genetic distance, 0.00036–0.00065), indicating a close phylogenetic relationship.

Genetic Variation

WGS-based genetic variation analysis was conducted across HAdV-14p1 subclades I–III using the HAdV-14 prototype strain as the reference genome (Supplementary Figure S2, available at <https://weekly.chinacdc.cn/>). Comparative genomic analysis revealed that the three subclades shared 99 specific nucleotide variations, comprising 85 single-nucleotide substitutions, six insertions, and eight deletions. These variations were distributed across the entire genome, whereas insertions and deletions were predominantly located in regions 11,000 bp upstream and 31,500 bp downstream. Subclade-specific variations were identified: subclade I harbored four substitutions and one insertion; subclade II contained six substitutions, one insertion, and one deletion; and subclade III possessed three nucleotide substitutions.

Detailed analysis of the 12 key functional regions identified 28 shared nucleotide variations across all three HAdV-14p1 subclades. These variations were distributed across 10 protein-coding regions (excluding pIX and pX), resulting in 15 amino acid changes (Figure 2). A shared six-nucleotide deletion

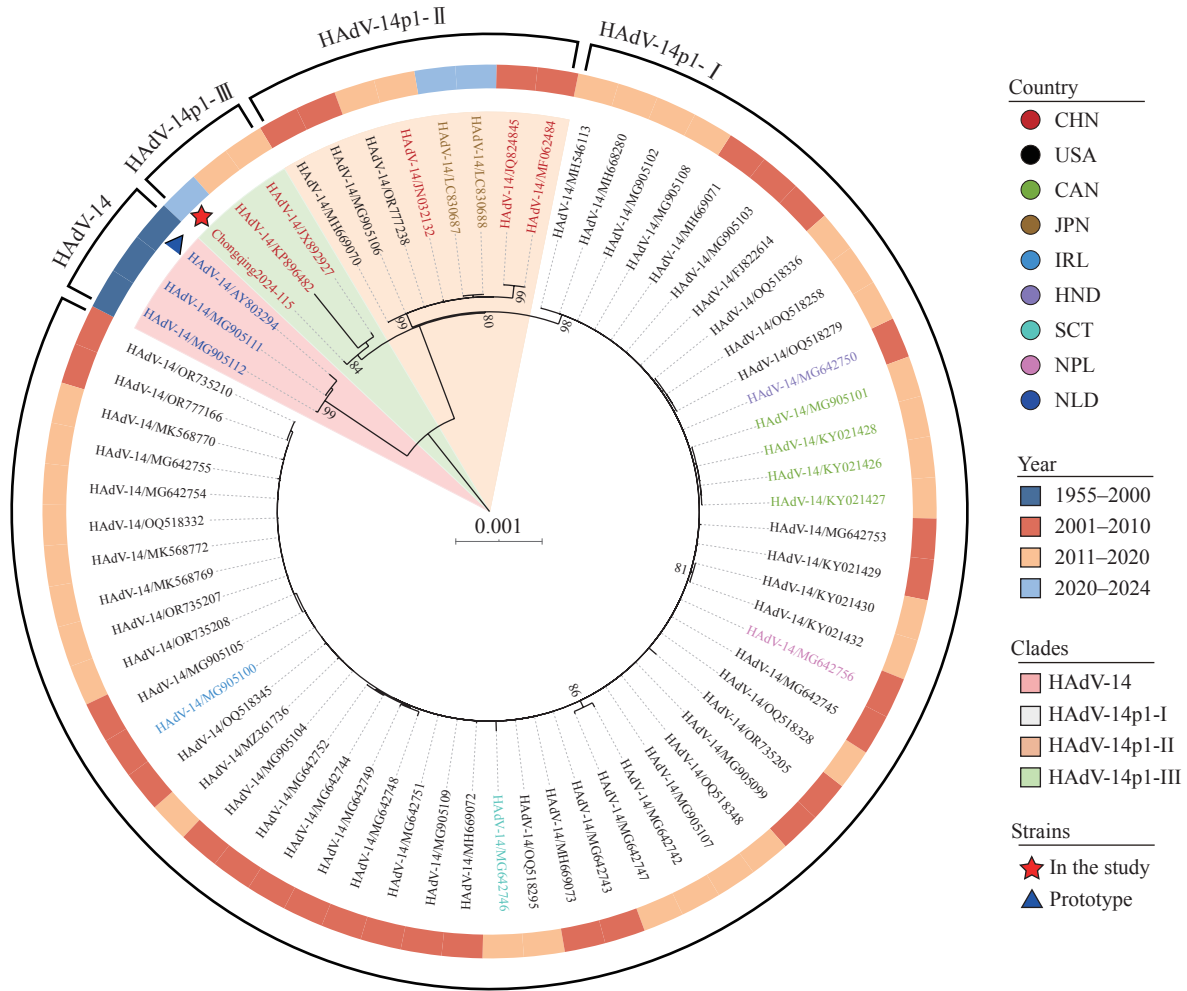


FIGURE 1. Maximum-likelihood phylogenetic tree of 68 HAdV-14 whole-genome sequences, including strain Chongqing2024-115 in this study and 67 strains from GenBank database. Abbreviation: HAdV-14=Human adenovirus type 14.

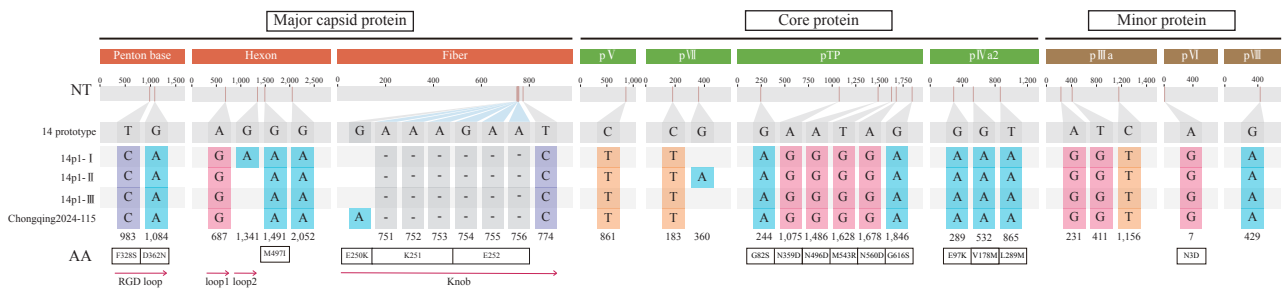


FIGURE 2. Nucleotide and amino acid variations specific to HAdV-14p1 clades and strain Chongqing2024-115 in 10 key coding regions compared to the HAdV-14 prototype strain (de Wit, AY803294). Note: In the schematic representation, major capsid proteins are shown in orange, core proteins in green, and minor proteins in brown. The annotation “NT” denotes nucleotide variation. “-” indicates nucleotide deletion, and the “AA” row displays amino acid variations resulting from mutations. Important functional regions are marked with red arrows.

(nt751–756) within the fiber gene, located in the knob domain, resulted in the loss of two amino acids (K251 and E252). Two shared nonsynonymous variations in the penton base gene (T983C and G1084A), located

within the RGD loop region, caused amino acid substitutions F328S and D362N, respectively. Subclade-specific nucleotide mutations were detected, including a synonymous mutation in the hexon gene

(G1341A) specific to subclade I and a synonymous mutation in the pVII gene (G360A) unique to subclade II. Chongqing2024-115 was characterized by two unique nucleotide substitutions: a synonymous mutation (T5601C) in the DNA polymerase gene and a nonsynonymous mutation (G31517A; E250K) within the knob region of the fiber gene (Supplementary Figure S2).

Genetic Recombination

A comprehensive recombination analysis was performed on all available HAdV-14p1 strains, including Chongqing2024-115, using WGSs from nine species B prototype strains retrieved from GenBank (recombinant strains were excluded to avoid analytical bias). The results indicated that no intra-species recombination events were detectable within the HAdV-14p1 clade.

DISCUSSION

Since 2019, no cases of HAdV-14p1 infection have been reported in China. As a highly contagious pathogen associated with substantial hospitalization and mortality, understanding the genetic origin of the recently identified HAdV-14p1 is crucial. We performed a comprehensive genomic characterization of the HAdV-14 strain Chongqing2024-115, isolated from a pediatric patient hospitalized with bronchopneumonia. Phylogenetic analysis revealed that this strain clustered closely with earlier HAdV-14p1 subclade III isolates obtained from respiratory infection outbreaks in Beijing (2012) and Gansu (2013) (13). The low intra-subclade genetic distance (0.00065) further supports a common evolutionary origin among these viruses, suggesting sustained transmission of this subclade within China. The absence of reported cases over the past decade may indicate prolonged cryptic transmission of this virus in the population, potentially due to factors such as insufficient surveillance efforts or the failure to systematically capture cases with mild symptoms, leading to a detection gap.

Compared with subclade III, the other two subclades (subclade I, detected across six countries during 2003–2020; subclade II, identified in three countries between 2006 and 2024) demonstrated broader spatiotemporal distributions. These subclades have maintained considerable genomic conservation over nearly two decades, as reflected by their low intra-

subclade genetic distance (≤ 0.00065). All three subclades exhibited high sequence identity (99.5%–99.6%) with the prototype HAdV-14 strain (de Wit, 1955), and the 99 shared variations (85 substitutions, six insertions, and eight deletions) represent a conserved genetic signature of the HAdV-14p1 variant that has circulated globally since its re-emergence after an approximately 50-year absence. Notably, the six-nucleotide deletion in the fiber gene (nt751–756) resulted in a two-amino acid deletion (K251 and E252) in the knob domain. This deletion has previously been identified as a hallmark genetic characteristic distinguishing the HAdV-14p1 strain from the HAdV-14 prototype (14). This two-amino acid deletion has been consistently present in all globally circulating strains identified since 2003, suggesting an important role in the adaptive evolution of HAdV-14p1. Additionally, strain Chongqing2024-115 harbors a unique amino acid substitution (E250K) within the knob domain of the fiber gene. Given the critical role of this domain in mediating viral attachment to and entry into host cells, further investigation is needed to determine whether this substitution affects viral infectivity.

The subdivision of HAdV-14p1 into three distinct subclades underscores the importance of high-resolution genomic data in tracking pathogen evolution. Although the overall genetic distances between subclades were minimal (0.00036–0.00065), consistent clustering patterns suggest that subtle genetic changes may be associated with adaptive evolution or host-specific interactions. The identification of subclade-specific variations and strain-specific mutations, such as those observed in the Chongqing2024-115 isolate, indicates ongoing microevolution within the HAdV-14p1 clade. Despite the detection of subclades II and III in China since 2011, research on HAdV-14p1 remains limited, with only five relevant Chinese genomic sequences (2010–2013) available in public databases. This scarcity of genetic data constrains a comprehensive understanding of the molecular epidemiology and endemic transmission patterns of HAdV-14p1 in China. Therefore, strengthening genomic surveillance is necessary to elucidate the prevalence, evolution, and public health impact of HAdV-14p1 in China.

In conclusion, WGS-based analysis provided a detailed view of the genetic characteristics of the newly identified HAdV-14p1 strain in China in 2024. Our findings confirm that the contemporary HAdV-14p1 strain shares common genetic ancestry with earlier

circulating Chinese subclade III strains. Given the association between HAdV-14p1 and severe illness, ongoing surveillance is imperative to gain a deeper understanding of its prevalence and evolution in China and to provide a basis for the formulation of targeted prevention and control measures.

Acknowledgments: The sentinel surveillance hospital in Chongqing for providing the viral strains and associated clinical data. This study used only viral strains and did not involve specimen collection.

Conflicts of interest: No conflicts of interest.

Funding: Supported by the National Key R&D Program of China (2022YFC2305303).

doi: 10.46234/ccdcw2026.033

* Corresponding authors: Zhen Zhu, zhuzhen@ivdc.chinacdc.cn; Hongmei Xu, xuhongm0095@hospital.cqmu.edu.cn.

¹ National Key Laboratory of Intelligent Tracking and Forecasting for Infectious Disease, NHC Key Laboratory of Medical Virology and Viral Diseases, National Institute for Viral Disease Control and Prevention, Chinese Center for Disease Control and Prevention, Beijing, China; ² Department of Infectious Diseases, Children's Hospital Affiliated to Chongqing Medical University, Chongqing, China.

Copyright © 2026 by Chinese Center for Disease Control and Prevention. All content is distributed under a Creative Commons Attribution Non Commercial License 4.0 (CC BY-NC).

Submitted: November 03, 2025

Accepted: January 19, 2026

Issued: February 20, 2026

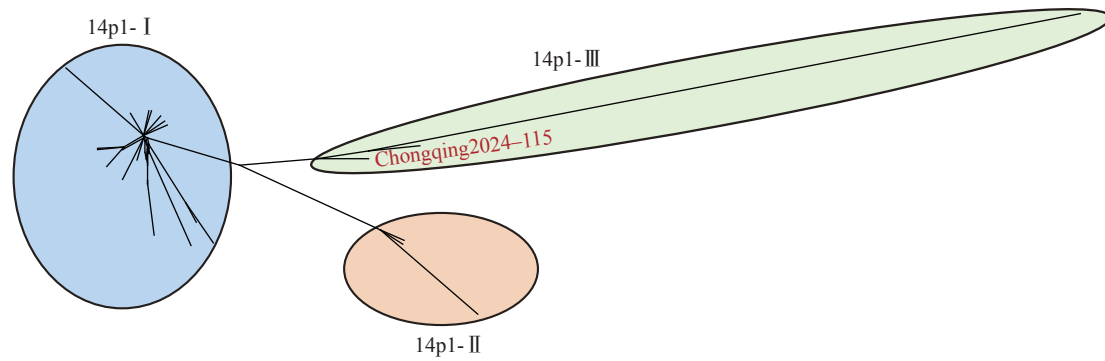
REFERENCES

1. Davison AJ, Benkő M, Harrach B. Genetic content and evolution of adenoviruses. *J Gen Virol* 2003;84(Pt 11):2895-908. <http://dx.doi.org/10.1099/vir.0.19497-0>.
2. Mao NY, Zhu Z, Zhang Y, Xu WB. Current status of human adenovirus infection in China. *World J Pediatr* 2022;18(8):533 – 7. <https://doi.org/10.1007/s12519-022-00568-8>.
3. Liu MC, Xu Q, Li TT, Wang T, Jiang BG, Lv CL, et al. Prevalence of human infection with respiratory adenovirus in China: a systematic review and meta-analysis. *PLoS Negl Trop Dis* 2023;17(2):e0011151. <https://doi.org/10.1371/journal.pntd.0011151>.
4. Van Der Veen J, Kok G. Isolation and typing of adenoviruses recovered from military recruits with acute respiratory disease in The Netherlands. *Am J Hyg* 1957;65(2):119 – 29. <https://doi.org/10.1093/oxfordjournals.aje.a119860>.
5. Lewis PF, Schmidt MA, Lu XY, Erdman DD, Campbell M, Thomas A, et al. A community-based outbreak of severe respiratory illness caused by human adenovirus serotype 14. *J Infect Dis* 2009;199(10):1427 – 34. <https://doi.org/10.1086/598521>.
6. O'Flanagan D, O'Donnell J, Domegan L, Fitzpatrick F, Connell J, Coughlan S, et al. First reported cases of human adenovirus serotype 14p1 infection, Ireland, October 2009 to July 2010. *Euro Surveill* 2011;16(8):19801. <https://pubmed.ncbi.nlm.nih.gov/21371411/>.
7. Girouard G, Garceau R, Thibault L, Oussedik Y, Bastien N, Li Y. Adenovirus serotype 14 infection, New Brunswick, Canada, 2011. *Emerg Infect Dis* 2013;19(1):119 – 22. <https://doi.org/10.3201/eid1901.120423>.
8. Parcell BJ, McIntyre PG, Yirell DL, Fraser A, Quinn M, Templeton K, et al. Prison and community outbreak of severe respiratory infection due to adenovirus type 14p1 in Tayside, UK. *J Public Health (Oxf)* 2015;37(1):64 – 9. <https://doi.org/10.1093/pubmed/fdu009>.
9. Mizuno S, Tanimoto Y, Mori A, Fuseya T, Ishida Y, Nishiyama M, et al. Acute encephalopathy associated with human adenovirus type 14 infection in 7-year-old girl, Japan. *Emerg Infect Dis* 2025;31(2):377 – 9. <https://doi.org/10.3201/eid3102.241168>.
10. Carr MJ, Kajon AE, Lu XY, Dunford L, O'Reilly P, Holder P, et al. Deaths associated with human adenovirus-14p1 infections, Europe, 2009-2010. *Emerg Infect Dis* 2011;17(8):1402 – 8. <https://doi.org/10.3201/eid1708.101760>.
11. Zhang QW, Seto D, Zhao SH, Zhu L, Zhao W, Wan CS. Genome sequence of the first human adenovirus type 14 isolated in China. *J Virol* 2012;86(12):7019 – 20. <https://doi.org/10.1128/jvi.00814-12>.
12. Huang GH, Yu DS, Zhu Z, Zhao H, Wang P, Gray GC, et al. Outbreak of febrile respiratory illness associated with human adenovirus type 14p1 in Gansu Province, China. *Influenza Other Respir Viruses* 2013;7(6):1048 – 54. <https://doi.org/10.1111/irv.12118>.
13. Mi ZQ, Butt AM, An XP, Jiang T, Liu W, Qin CF, et al. Genomic analysis of HAdV-B14 isolate from the outbreak of febrile respiratory infection in China. *Genomics* 2013;102(5-6):448 – 55. <https://doi.org/10.1016/j.ygeno.2013.09.001>.
14. Liao LN, Chen YH, Wang YY, Ren RW, Li J, Zhang S, et al. Genomic characteristics and phylogenetic analysis of human adenovirus 14 in China during 2010-2016. *Infect Genet Evol* 2025;134:105805. <https://doi.org/10.1016/j.meegid.2025.105805>.
15. Zhu WW, Wu WW, Xu YP, Du YG, Tang LY, Tong J. Epidemiological characteristics and genotypes of human adenovirus among children in Xuzhou from 2013 to 2017. *Int J Virol* 2018;25(5):315 – 8. <https://doi.org/10.3760/cma.j.issn.1673-4092.2018.05.008>.

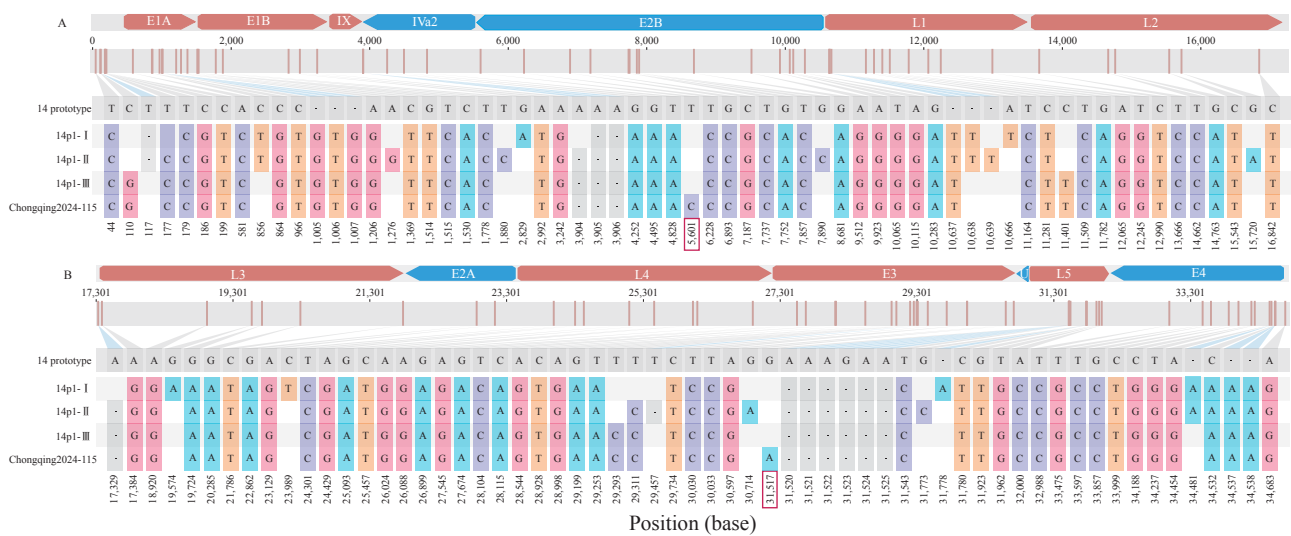
SUPPLEMENTARY MATERIAL

SUPPLEMENTARY TABLE S1. Genomic annotations of strain Chongqing2024-115.

Gene	Encoded product	Chongqing2024-115
E1A	29.1 kDa protein	563-1,142; 1,227-1,435
	6.5 kDa protein	563-634; 1,227-1,331
E1B	20 kDa protein	1,605-2,147
	54.9 kDa protein	1,910-3,394
IX	pIX protein	3,477-3,896
IVa2	IVa2 protein	3,963-5,296; 5,575-5,587
E2B	DNA polymerase	5,066-8,638; 13,625-13,633
	terminal protein precursor	8,437-10,398; 13,625-13,633
L1	43 kDa protein	10,649-11,809
	protein IIIa precursor	11,835-13,599
L2	penton protein	13,681-15,357
	protein VII	15,362-15,940
	protein V precursor	15,983-17,038
	protein X	17,067-17,297
	protein VI	17,378-18,118
L3	hexon protein	18,234-21,071
	23 kDa protein	21,108-21,737
E2A	DNA binding protein	21,815-23,371
L4	100 kDa hexon-assembly associated protein	23,402-25,840
	33 kDa protein	25,572-25,890; 26,060-26,421
	22 kDa protein	25,572-26,147
	protein VIII	26,471-27,154
E3	11.7 kDa protein	27,154-27,471
	14.6 kDa protein	27,425-27,820
	18.4 kDa protein	27,805-28,305
	20.1 kDa protein	28,325-28,870
	20.8 kDa protein	28,888-29,439
	10.1 kDa protein	29,483-29,758
	14.9 kDa protein	29,763-30,167
	15 kDa protein	30,160-30,567
U	U protein	30,591-30,755
L5	fiber protein	30,770-31,741
E4	Orf6/7 protein	31,777-32,028; 32,751-32,924
	Orf6 protein	32,025-32,924
	Orf4 protein	32,827-33,195
	Orf3 protein	33,204-33,557
	Orf2 protein	33,554-33,943
	Orf1 protein	33,986-34,363



SUPPLEMENTARY FIGURE S1. Phylogenetic network generated based on WGS of 65 HAdV-14p1 strains. Abbreviation: WGS=Whole Genome Sequence; HAdV-14=Human adenovirus type 14.



SUPPLEMENTARY FIGURE S2. Nucleotide variations specific to HAdV-14p1 clades and strain Chongqing2024-115 in the whole genome compared to the HAdV-14 prototype strain (de Wit, AY803294). (A) The genomic fragment 1–17,300 bp; (B) The fragment 17,301 bp to the end.

Notes: “-” indicates nucleotide deletion; variation sites unique to the strain Chongqing 2024-115 are highlighted with red boxes.