

Perspectives

Advancements in Defining and Estimating the Reproduction Number in Infectious Disease Epidemiology

Kangguo Li^{1,2}; Jiayi Wang^{1,2}; Jiayuan Xie^{1,2}; Jia Rui^{1,2}; Buasiyamu Abudunaibi^{1,2}; Hongjie Wei^{1,2};
Hong Liu^{1,2}; Shuo Zhang^{1,2}; Qun Li³; Yan Niu^{3,4}; Tianmu Chen^{1,2,5}

The reproduction number (R) serves as a fundamental metric in the examination of infectious disease outbreaks, epidemics, and pandemics. Despite an array of available methods for estimating R , both newcomers and established public health professionals often encounter difficulties in comprehending the circumstances for their use and their constrictions. Consequently, this review intends to offer elementary guidance on R 's selection and estimation approaches. To facilitate our review, we executed an extensive search on PubMed and Web of Science applying the following search approach: ["Basic Reproduction Number/classification"(Mesh)] AND ["Basic Reproduction Number/prevention and control"(Mesh)] OR ["Basic Reproduction Number/statistics and numerical data"(Mesh)]. Our search parameters were restricted to articles published from January 2013 to January 2023. This search rendered a total of 7,094 articles, of which we selected 60 that met our inclusion standards for further analysis.

CONCEPTUAL UNDERSTANDING OF R : AN ANALYSIS

R is a fundamental measure that indicates the average number of infections or cases resulting from contact with an infected individual, thus serving as an important gauge of the transmissibility of infectious diseases. There are three types of R : basic reproduction number (R_0), effective reproduction number (R_{eff}), and real-time or time-varying reproduction number (R_t). R_0 is utilized for evaluating the transmissibility of new pathogens or variants when they emerge (1). However, R_{eff} and R_t are employed to assess the effectiveness of public health and social measures (PHSMs), providing valuable insights for policymakers and public health officials (Figure 1) (2–4).

R_0 , also known as the basic reproduction number, signifies the mean number of secondary infections attributed solely to a single infected individual within a

susceptible population (5–7). It proves instrumental in predicting the probability and magnitude of disease outbreaks, plus the vaccination threshold required to establish herd immunity (1,8). Various factors like the frequency of contact among the population, sanitary practices, and seasonal changes may alter R_0 further (9). Altering the transmission rate (β), the recovery rate (γ or inverse of the mean infection period), or the contact rate substantially influences the estimated value of R_0 (10). It is essential to account for any pre-existing immunity within the given population while calculating R_0 . Presently, there exists no standardized method for determining and reporting R_0 , addressing the issue of its variability (11).

The concept of R_{eff} is similar to R_0 and often confused by researchers. The major distinction lies in the fact that R_{eff} is suitable for establishing a baseline for PHSMs or exposed populations, representing the actual immunity of the population (12). As a result, R_{eff} is usually smaller than R_0 , because it primarily relies on not only the transmissibility of pathogens but also the levels of immunization within the population (13).

Anne Cori et al. (14) provided a more detailed breakdown of R_t , dividing it into the case reproduction number (R_c) and the instantaneous reproduction number (R_i). R_c represents the average number within R_t and reflects the transmissibility at a given time point. On the other hand, R_i represents the average number within R_t calculated under the assumption of no change after a specific time point, making it easier to estimate in real-time (15). R_t estimates the spread of pathogens by monitoring and tracking data that evolves over time during the course of an outbreak (16). R_t is also an important parameter for describing the epidemiological characteristics of a disease and evaluating the effectiveness of PHSMs (Figure 1) (17–18). The values of R_t vary due to factors such as changes in immunity and interventions across different populations, including interventions that impact personal contact networks (19–20). In practice, researchers must choose whether the main R_t index to














Application scenarios		Direct	Indirect			
		DBM	DBM	NGM	FSE	GIBM
R_0	 Probability size & final size Needed vaccination coverage					
R_{eff}	 Probability size & final size Policy formulation Intervention effectiveness					
R_t	 Change in transmission Policy formulation Intervention effectiveness					

FIGURE 1. Comparison of application scenarios for various reproduction number methods.

Abbreviation: DBM=definition-based method; NGM=next-generation method; FSE=final-size equation; GIBM=generation interval-based method.

be obtained is R_c or R_i , and then select the appropriate modeling methods accordingly. Overall, both R_c and R_i represent the average number of individuals who are at risk of infection at a specific time (t), with R_c focusing on the attributes of infected individuals at the time t and being more widely used, while R_i emphasizes the temporal attribute at time t if the situation remains unchanged. Consequently, if the disease transmissibility declines at a particular point, R_i will transition from high to low, while R_c will smoothly decrease (21).

METHODOLOGY FOR CALCULATING R

Implementation of the Direct Method:

The direct method is used to estimate R by analyzing a clear transmission chain multiplying the β with the transmission probability per contact (p), contact rate (c), and infectious period (D) (11,22):

$$\beta = pc$$

$$R_0 = \beta D = pcD$$

The direct method is applicable to distinct scenarios that involve a minimal number of case generations within a brief time frame, or small sample sizes during the early phase of an epidemic or outbreak. This allows researchers the potential to separately calculate R for each possible transmission chain, analyze the

distribution of R , and evaluate the contributions of different transmission chains to the spread of the disease. However, the direct method might be prone to bias resulting from small sample sizes and is subject to limitations related to the lack of time variation. Moreover, challenges regarding underreporting and fragmented data in real-time evaluations present potential issues (23).

Implementation of the Indirect Method

Methodology Based on Definitions: The definition-based method (DBM) is an indirect approach used to estimate the R value. This method is applied to various transmission dynamics models, including the Susceptible-Infectious-Recovered (SIR) model, the Susceptible-Exposed-Infectious-Recovered (SEIR) model, the Susceptible-Infectious-Recovered-Cross immune (SIRC) model, and the Susceptible-Infectious-Recovered-Susceptible (SEIS) model (24–28). Taking the SIR model as an example:

$$\frac{dS}{dt} = b_r N - \frac{\beta SI}{N} - d_r S$$

$$\frac{dI}{dt} = \frac{\beta SI}{N} - \gamma I - d_r I$$

$$\frac{dR}{dt} = \gamma I - d_r R$$

The secondary infections generated by an infected individual per unit of time are represented as $\beta S/N$,

which corresponds to the inflow process. On the other hand, the recovery or natural death of an infected individual per unit of time is denoted as $\gamma + d_r$, which corresponds to the outflow process. Thus, we can calculate R_{eff} as follows:

$$R_{eff} = \frac{\text{Inflow process}}{\text{Outflow process}} = \frac{\beta S}{N} \times \frac{1}{\gamma + d_r} = \frac{\beta S}{(\gamma + d_r)N}$$

R_0 refers to the R when nearly the entire population is susceptible, which means S is approximately equal to N :

$$R_0 = \frac{\beta}{d_r + \gamma}$$

The DBM calculates R by expressing it as a function of model parameters. This approach proves valuable in the advanced stages of an epidemic as it yields results with significant explanatory power. However, its applicability is limited to single-host and single-kinetic models, thus restricting its use in multi-host or co-kinetic models. The DBM incorporates both the disease's natural history and demographic parameters, rendering it meaningful for predicting and preventing outbreaks. Moreover, it is renowned for its simplicity, ease of comprehension, and minimal hardware or software requirements.

Methodology Based on Next-Generation: The next-generation method (NGM) serves as a prevalent approach for the estimation of R . This method utilizes the maximum eigenvalue of the next-generation matrix within a dynamic model following the method proposed by Van den Driessche and Watmough (29–33). NGM is frequently applied across a range of dynamic models including, but not limited to, the SIR and SEIS models (25). Furthermore, it delivers quantitative accounts of secondary infections and can estimate the percentage of undetected cases across diverse outbreak scenarios (29,34). Compartments within these dynamic models are differentiated based on their infectivity. The ‘x-group’ signifies compartments possessing infectivity, whereas the ‘y-group’ denotes compartments devoid of infectivity. The equations corresponding to these groups are presented below:

$$\frac{dx_i}{dt} = F_i(x, y) - V_i(x, y) \quad i = 1, \dots, n$$

$$\frac{dy_j}{dt} = G_j(x, y) \quad j = 1, \dots, m$$

F_i represents the newly infected individuals in compartment i , V_i represents individuals who transit to other compartments. To illustrate NGM, we will

continue using the SIR model as an example. In the SIR model, where n and m are 1 and 2, respectively, and with $x = I$ and $y = (S, R)$, the corresponding equations are as follows:

$$F_1 = \frac{\beta SI}{N}$$

$$V_1 = \gamma I + d_r I$$

$$G_1 = b_r N - \frac{\beta SI}{N} - d_r S$$

$$G_2 = \gamma I - d_r R$$

Taking derivatives of F and V to I , one obtains the Jacobi matrix: $F = \beta S/N$, and $V = \gamma + d_r$. And R_{eff} is the real part of the leading eigenvalue of the next-generation matrix (FV^{-1}) 25:

$$R_{eff} = \rho(FV^{-1}) = \frac{F}{V} = \frac{\frac{\beta S}{N}}{\gamma + d_r} = \frac{\beta S}{(\gamma + d_r)N}$$

$$R_0 = \frac{\beta}{\gamma + d_r}$$

Nevertheless, the application of the NGM method to multi-group or multi-host compartmental models exhibits certain limitations. This method exclusively ascertains the stability threshold of a disease-free equilibrium, displaying a deficiency in explicit explanatory power. Employing smaller data sets during the initial phases of an epidemic may result in the omission of pivotal information. Over time, there has been a noted enhancement in the quality and dependability of the NGM results. Hence, researchers must modify their methodologies based on specific scenarios. For instance, when studying diseases such as hand, foot, and mouth disease, it might be plausible to exclude certain factors like the short disease duration, mobility of patients, and spatial structure.

Equation for Determining Final Size: The final-size equation (FSE) is a valuable tool for comprehending the relationship between the outcome of an epidemic and R_0 , while taking into account the proportions of susceptible and recovered individuals. In the SIR model, the calculation formula is as follows:

$$R_0 = \frac{\ln \frac{S_0}{S_\infty}}{1 - S_\infty}$$

Where S_0 and S_∞ represent the initial and final proportions of susceptible individuals.

FSE is often employed in the SIR model to ascertain the ultimate scale of an epidemic (35). With its precise data output and straightforward equation form, it is well suited to facilitate initial estimates following the conclusion of an epidemic. Nonetheless, its use is

model-specific and necessitates fresh derivation for application for other models, which can prove challenging for complex dynamic models.

It has been definitively established that the FSE possesses a unique solution in three mean field models, namely homogeneous, pairwise, and heterogeneous. Moreover, linearizing the FSE facilitates the transformation of optimal vaccination issues into simpler knapsack problems, yielding practical insights for decision-makers and the general public when considering vaccination strategies (36–37). However, a gap exists with respect to the availability of an R package incorporating displacement or interaction for the calculation of R_t using the FSE approach (38).

Methodology Based on Generation Intervals: The method based on generation interval is frequently utilized to estimate R_t in the field of epidemiology. This approach leverages the concept of the generation gap, defined as the duration between the infection of a primary case and the consequent infection of secondary cases. This method streamlines the natural history of the illness by concentrating on the distribution of time intervals among generations. Within this framework, two key indicators are emphasized: the generation interval (GT) and the serial interval (SI). GT signifies the duration between infection incidents in an infector-infected pair, whereas SI symbolizes the time from symptom onset in these pairs (39). Accurate estimation of GT becomes demanding as it is dependent on an exhaustive investigation of contact history (40). In comparison, SI's determination is less challenging as symptoms can be readily detected during field epidemiological surveys (41). By quantifying the relationship between generations using SI, researchers can estimate R_t , R_{eff} , and R_0 (42–44).

Several R (version 4.3.0, R Core Team, Vienna Austria) packages, namely EpiEstim, EpiNow2, and R0, currently facilitate the computation of regeneration numbers based on GT or SI (15,45–46), thereby significantly lowering the barrier to their utilization. We have developed an interactive application for users unfamiliar with the R language, particularly grassroots disease control staff. This application, called Reproduction Number Calculator, enables access to these R packages without necessitating knowledge of programming (available at <https://toolbox.ctmodelling.cn/>). However, it is crucial to acknowledge the method's inherent limitations. Inaccuracies may arise if the assumed distribution of intergenerational times does not accurately reflect the dynamics of the disease (42). This uncertainty in

distribution can potentially result in an underestimation of R 's uncertainty (15). Oversights related to group immunity and infection staging can create bias when estimating R_{eff} (42). Further, the generation interval-based method comes with specific demands and limitations, such as a need for clear transmission chains, comprehensive and timely data, and an accurate intergenerational time distribution assumption. These factors may limit its utility in certain scenarios.

In conclusion, the generation interval-based method provides valuable insights into disease transmission dynamics and facilitates the estimation of R_t , R_{eff} and R_0 . However, researchers should exercise caution in interpreting the results and consider the assumptions and data requirements associated with the method.

CONCLUSION

Choosing the correct approach to R estimation is critical in epidemiological research. Each model introduces its own unique strengths and weaknesses. The desired R , dictated by disease characteristics and accessible data, must be thoughtfully considered by researchers to identify the most fitting calculation method. This systematic strategy guarantees that the estimation procedure corresponds with the existing conditions and provides trustworthy outcomes.

Conflicts of interest: No conflicts of interest.

Funding: Supported by the National Key R&D Program of China (2021ZD0113903, 2021YFC2301604), Fundamental Research Funds for the Central Universities (20720230001).

doi: 10.46234/ccdcw2023.158

Corresponding authors: Tianmu Chen, chentianmu@xmu.edu.cn; Yan Niu, niuyan@chinacdc.cn.

¹ State Key Laboratory of Vaccines for Infectious Diseases, Xiang An Biomedicine Laboratory, School of Public Health, Xiamen University, Xiamen City, Fujian Province, China; ² State Key Laboratory of Molecular Vaccinology and Molecular Diagnostics, National Innovation Platform for Industry-Education Integration in Vaccine Research, Xiamen University, Xiamen City, Fujian Province, China; ³ Chinese Center for Disease Control and Prevention, Beijing, China.

Submitted: July 18, 2023; Accepted: September 11, 2023

REFERENCES

1. Dhungel B, Rahman S, Rahman M, Bhandari AKC, Le PM, Biva NA, et al. Reliability of early estimates of the basic reproduction number of COVID-19: a systematic review and meta-analysis. *Int J Environ Res Public Health* 2022;19(18):11613. <http://dx.doi.org/10.3390/IJERPH191811613>.

2. Gostic KM, McGough L, Baskerville EB, Abbott S, Joshi K, Tedijanto C, et al. Practical considerations for measuring the effective reproductive number, R_e . *PLoS Comput Biol* 2020;16(12):e1008409. <http://dx.doi.org/10.1371/journal.pcbi.1008409>.
3. Ferguson NM, Donnelly CA, Anderson RM. Transmission intensity and impact of control policies on the foot and mouth epidemic in Great Britain. *Nature* 2001;413(6855):542–8. <http://dx.doi.org/10.1038/35097116>.
4. Linka K, Peirlinck M, Kuhl E. The reproduction number of COVID-19 and its correlation with public health interventions. *Comput Mech* 2020;66(4):1035–50. <http://dx.doi.org/10.1007/s00466-020-01880-8>.
5. Pijpers FP. A non-parametric method for determining epidemiological reproduction numbers. *J Math Biol* 2021;82(5):37. <http://dx.doi.org/10.1007/s00285-021-01590-6>.
6. Navarro Valencia VA, Díaz Y, Pascale JM, Boni MF, Sanchez-Galan JE. Using compartmental models and Particle Swarm Optimization to assess Dengue basic reproduction number R_0 for the Republic of Panama in the 1999-2022 period. *Heliyon* 2023;9(4):e15424. <http://dx.doi.org/10.1016/j.heliyon.2023.e15424>.
7. Takahashi S, Liao QH, Van Boeckel TP, Xing WJ, Sun JL, Hsiao VY, et al. Hand, foot, and mouth disease in China: modeling epidemic dynamics of enterovirus serotypes and implications for vaccination. *PLoS Med* 2016;13(2):e1001958. <http://dx.doi.org/10.1371/journal.pmed.1001958>.
8. Anderson RM, May RM. Infectious diseases of humans: dynamics and control. Oxford: Oxford University Press. 1991. <https://www.cambridge.org/core/services/aop-cambridge-core/content/view/S0950268800059896>.
9. Zhang XB, Yan DY, Chen C, Jiang DX, Ding C, Lan L, et al. Research progress on the basic and effective reproductive number in the epidemiology of infectious diseases. *Chin J Dis Control Prev* 2021;25(7):753-7, 790. <http://dx.doi.org/10.16462/j.cnki.zbjbkz.2021.07.003>. (In Chinese).
10. Ridenhour B, Kowalik JM, Shay DK. Unraveling R_0 : considerations for public health applications. *Am J Public Health* 2014;104(2):e32–41. <http://dx.doi.org/10.2105/AJPH.2013.301704>.
11. Guerra FM, Bolotin S, Lim G, Heffernan J, Deeks SL, Li Y, et al. The basic reproduction number (R_0) of measles: a systematic review. *Lancet Infect Dis* 2017;17(12):e420–8. [http://dx.doi.org/10.1016/S1473-3099\(17\)30307-9](http://dx.doi.org/10.1016/S1473-3099(17)30307-9).
12. Liu Y, Rocklöv J. The effective reproductive number of the Omicron variant of SARS-CoV-2 is several times relative to Delta. *J Travel Med* 2022;29(3):taac037. <http://dx.doi.org/10.1093/jtm/taac037>.
13. Green WD, Ferguson NM, Cori A. Inferring the reproduction number using the renewal equation in heterogeneous epidemics. *J Roy Soc Interface* 2022;19(188):20210429. <http://dx.doi.org/10.1098/rsif.2021.0429>.
14. Cori A, Ferguson NM, Fraser C, Cauchemez S. A new framework and software to estimate time-varying reproduction numbers during epidemics. *Am J Epidemiol* 2013;178(9):1505–12. <http://dx.doi.org/10.1093/aje/kwt133>.
15. Thompson RN, Stockwin JE, van Gaalen RD, Polonsky JA, Kamvar ZN, Demarsh PA, et al. Improved inference of time-varying reproduction numbers during infectious disease outbreaks. *Epidemics* 2019;29:100356. <http://dx.doi.org/10.1016/j.epidem.2019.100356>.
16. Nishiura H, Chowell G. The effective reproduction number as a prelude to statistical estimation of time-dependent epidemic trends. In: Chowell G, Hyman JM, Bettencourt LMA, Castillo-Chavez C, editors. Mathematical and statistical estimation approaches in epidemiology. Dordrecht: Springer. 2009. http://dx.doi.org/10.1007/978-90-481-2313-1_5.
17. Gressani O, Wallinga J, Althaus CL, Hens N, Faes C. EpiLPS: a fast and flexible Bayesian tool for estimation of the time-varying reproduction number. *PLoS Comput Biol* 2022;18(10):e1010618. <http://dx.doi.org/10.1371/journal.pcbi.1010618>.
18. Lal R, Huang WD, Li ZQ, Prasad S. An assessment of transmission dynamics via time-varying reproduction number of the second wave of the COVID-19 epidemic in Fiji. *Roy Soc Open Sci* 2022;9(8):220004. <http://dx.doi.org/10.1098/rsos.220004>.
19. Creswell R, Augustin D, Bouros I, Farm HJ, Miao S, Ahern A, et al. Heterogeneity in the onwards transmission risk between local and imported cases affects practical estimates of the time-dependent reproduction number. *Philos Trans Roy Soc A Math Phys Eng Sci* 2022;380(2233):20210308. <http://dx.doi.org/10.1098/RSTA.2021.0308>.
20. Eales O, Ainslie KEC, Walters CE, Wang HW, Archison C, Ashby D, et al. Appropriately smoothing prevalence data to inform estimates of growth rate and reproduction number. *Epidemics* 2022;40:100604. <http://dx.doi.org/10.1016/j.epidem.2022.100604>.
21. Fraser C. Estimating individual and household reproduction numbers in an emerging epidemic. *PLoS One* 2007;2(8):e758. <http://dx.doi.org/10.1371/journal.pone.0000758>.
22. Dietz K. The estimation of the basic reproduction number for infectious diseases. *Stat Methods Med Res* 1993;2(1):23–41. <http://dx.doi.org/10.1177/096228029300200103>.
23. Gao DZ, Lou YJ, He DH, Porco TC, Kuang Y, Chowell G, et al. Prevention and control of zika as a mosquito-borne and sexually transmitted disease: a mathematical modeling analysis. *Sci Rep* 2016;6:28070. <http://dx.doi.org/10.1038/srep28070>.
24. Martcheva M. An introduction to mathematical epidemiology. New York: Springer. 2015. <http://dx.doi.org/10.1007/978-1-4899-7612-3>.
25. Guo XH, Guo YC, Zhao ZY, Yang ST, Su YH, Zhao BH, et al. Computing R_0 of dynamic models by a definition-based method. *Infect Dis Model* 2022;7(2):196–210. <http://dx.doi.org/10.1016/j.idm.2022.05.004>.
26. He SB, Peng YX, Sun KH. SEIR modeling of the COVID-19 and its dynamics. *Nonlinear Dyn* 2020;101(3):1667–80. <http://dx.doi.org/10.1007/s11071-020-05743-y>.
27. Goel S, Bhatia SK, Tripathi JP, Bugalia S, Rana M, Bajjiya VP. SIRC epidemic model with cross-immunity and multiple time delays. *J Math Biol* 2023;87(3):42. <http://dx.doi.org/10.1007/S00285-023-01974-W>.
28. Jain S, Kumar S. Dynamic analysis of the role of innate immunity in SEIS epidemic model. *Eur Phys J Plus* 2021;136(4):439. <http://dx.doi.org/10.1140/epjp/s13360-021-01390-3>.
29. Tang B, Wang X, Li Q, Bragazzi NL, Tang SY, Xiao YN, et al. Estimation of the transmission risk of the 2019-nCoV and its implication for public health interventions. *J Clin Med* 2020;9(2):462. <http://dx.doi.org/10.3390/jcm9020462>.
30. Zhou WK, Wang AL, Xia F, Xiao YN, Tang SY. Effects of media reporting on mitigating spread of COVID-19 in the early phase of the outbreak. *Math Biosci Eng* 2020;17(3):2693–707. <http://dx.doi.org/10.3934/mbe.2020147>.
31. Yakob L, Clements ACA. A mathematical model of chikungunya dynamics and control: the major epidemic on Réunion Island. *PLoS One* 2013;8(3):e57448. <http://dx.doi.org/10.1371/journal.pone.0057448>.
32. Rudge JW, Webster JP, Lu DB, Wang TP, Fang GR, Basáñez MG. Identifying host species driving transmission of schistosomiasis japonica, a multihost parasite system, in China. *Proc Natl Acad Sci USA* 2013;110(28):11457–62. <http://dx.doi.org/10.1073/pnas.1221509110>.
33. Li Y, Zhang JH, Zhang XN. Modeling and preventive measures of hand, foot and mouth disease (HFMD) in China. *Int J Environ Res Public Health* 2014;11(3):3108–17. <http://dx.doi.org/10.3390/ijerph110303108>.
34. Unwin HJT, Cori A, Imai N, Gaythorpe KAM, Bhatia S, Cattarino L, et al. Using next generation matrices to estimate the proportion of infections that are not detected in an outbreak. *Epidemics* 2022;41:100637. <http://dx.doi.org/10.1016/j.epidem.2022.100637>.
35. Bidari S, Chen XY, Peters D, Pittman D, Simon PL. Solvability of implicit final size equations for SIR epidemic models. *Math Biosci* 2016;282:181–90. <http://dx.doi.org/10.1016/j.mbs.2016.10.012>.
36. Penn MJ, Donnelly CA. Asymptotic analysis of optimal vaccination policies. *Bull Math Biol* 2023;85(3):15. <http://dx.doi.org/10.1007/s11538-022-01114-3>.

37. Shaw CL, Kennedy DA. What the reproductive number R_0 can and cannot tell us about COVID-19 dynamics. *Theor Popul Biol* 2021;137:2 – 9. <http://dx.doi.org/10.1016/j.tpb.2020.12.003>.
38. Nash RK, Nouvellet P, Cori A. Real-time estimation of the epidemic reproduction number: scoping review of the applications and challenges. *PLoS Digit Health* 2022;1(6):e0000052. <http://dx.doi.org/10.1371/journal.pdig.0000052>.
39. Griffin J, Casey M, Collins Á, Hunt K, McEvoy D, Byrne A, et al. Rapid review of available evidence on the serial interval and generation time of COVID-19. *BMJ Open* 2020;10(11):e040263. <http://dx.doi.org/10.1136/bmjopen-2020-040263>.
40. Boëlle PY, Ansart S, Cori A, Valleron AJ. Transmission parameters of the A/H1N1 (2009) influenza virus pandemic: a review. *Influenza Other Respir Viruses* 2011;5(5):306 – 16. <http://dx.doi.org/10.1111/j.1750-2659.2011.00234.x>.
41. Svensson Å. A note on generation times in epidemic models. *Math Biosci* 2007;208(1):300 – 11. <http://dx.doi.org/10.1016/j.mbs.2006.10.010>.
42. Koh WM, Bogich T, Siegel K, Jin J, Chong EY, Tan CY, et al. The epidemiology of hand, foot and mouth disease in Asia: a systematic review and analysis. *Pediatr Infect Dis J* 2016;35(10):e285 – 300. <http://dx.doi.org/10.1097/INF.0000000000001242>.
43. Nouvellet P, Cori A, Garske T, Blake IM, Dorigatti I, Hinsley W, et al. A simple approach to measure transmissibility and forecast incidence. *Epidemics* 2018;22:29 – 35. <http://dx.doi.org/10.1016/j.epidem.2017.02.012>.
44. Townsend SE, Sumantra IP, Pudjiatmoko, Bagus GN, Brum E, Cleaveland S, et al. Designing programs for eliminating canine rabies from islands: Bali, Indonesia as a case study. *PLoS Negl Trop Dis* 2013;7(8):e2372. <http://dx.doi.org/10.1371/journal.pntd.0002372>.
45. Abbott S, Hickson J, Funk S, Badr HS, Monticone P, Ellis P, et al. epiforecasts/EpiNow2: 1.3.4 release. Zenodo; 2023. <http://dx.doi.org/10.5281/zenodo.7611804>.
46. Obadia T, Haneef R, Boëlle PY. The R0 package: a toolbox to estimate reproduction numbers for epidemic outbreaks. *BMC Med Inform Decis Mak* 2012;12:147. <http://dx.doi.org/10.1186/1472-6947-12-147>.